

Supporting Information

De Silva *et al.* 10.1073/pnas.0801993105

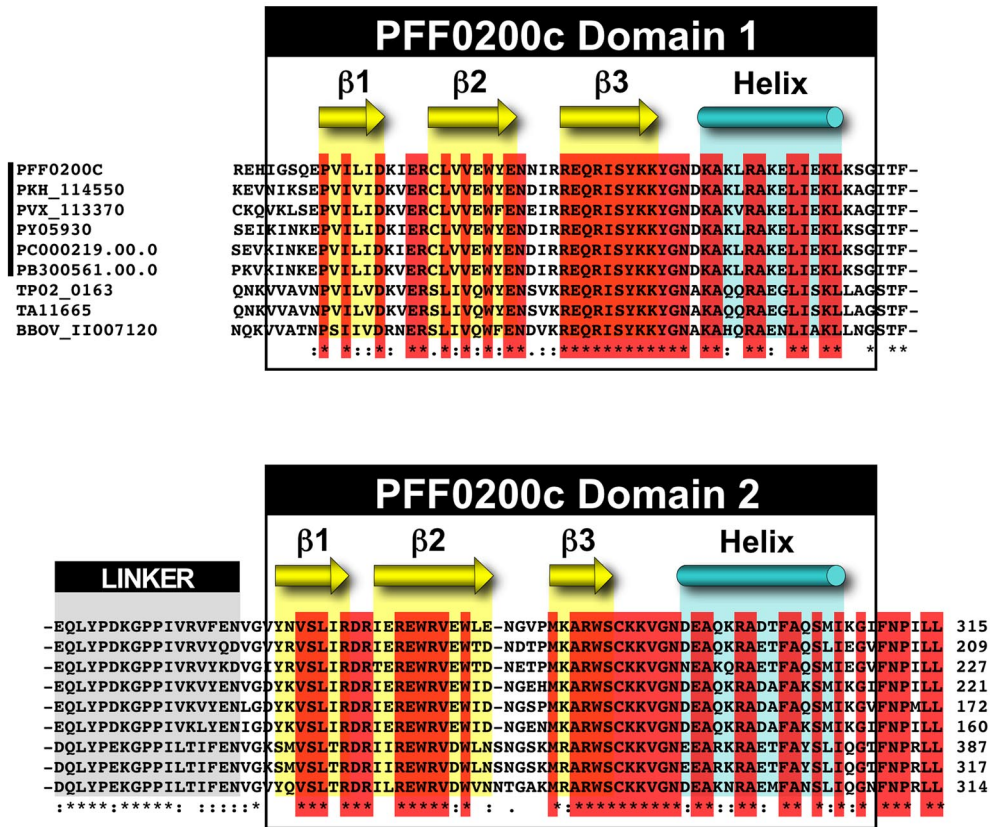


Fig. S1. Alignment of the AP2 domains (amino acids 177–311) from PFF0200c to orthologues in five additional *Plasmodium* spp. and three *Apicomplexan* species. The AP2 domain (boxed) is highly conserved across all species shown. Conservation of residues is most significant in the three β -strands (shaded yellow) of the AP2 domain, and is less significant in the α -helix (shaded blue). Absolutely conserved residues likely to be involved in DNA-binding are highlighted in red. Secondary structure predictions were predicted using Jnet [Cuff JA, Barton GJ (2000) Application of multiple sequence alignment profiles to improve protein secondary structure prediction. *Proteins* 40:502–511]. There are no orthologues detected in the other sequenced Apicomplexan species *T. gondii* and the *Cryptosporidia*. PF = *Plasmodium falciparum*, PVX = *P. vivax*, PKH = *P. knowlesi*, PB = *P. berghei*, PY = *P. yoelii*, PC = *P. chabaudi*, TP = *Theileria parvum*, TA = *Theileria annulata*, BBOV = *Babesia bovis*.

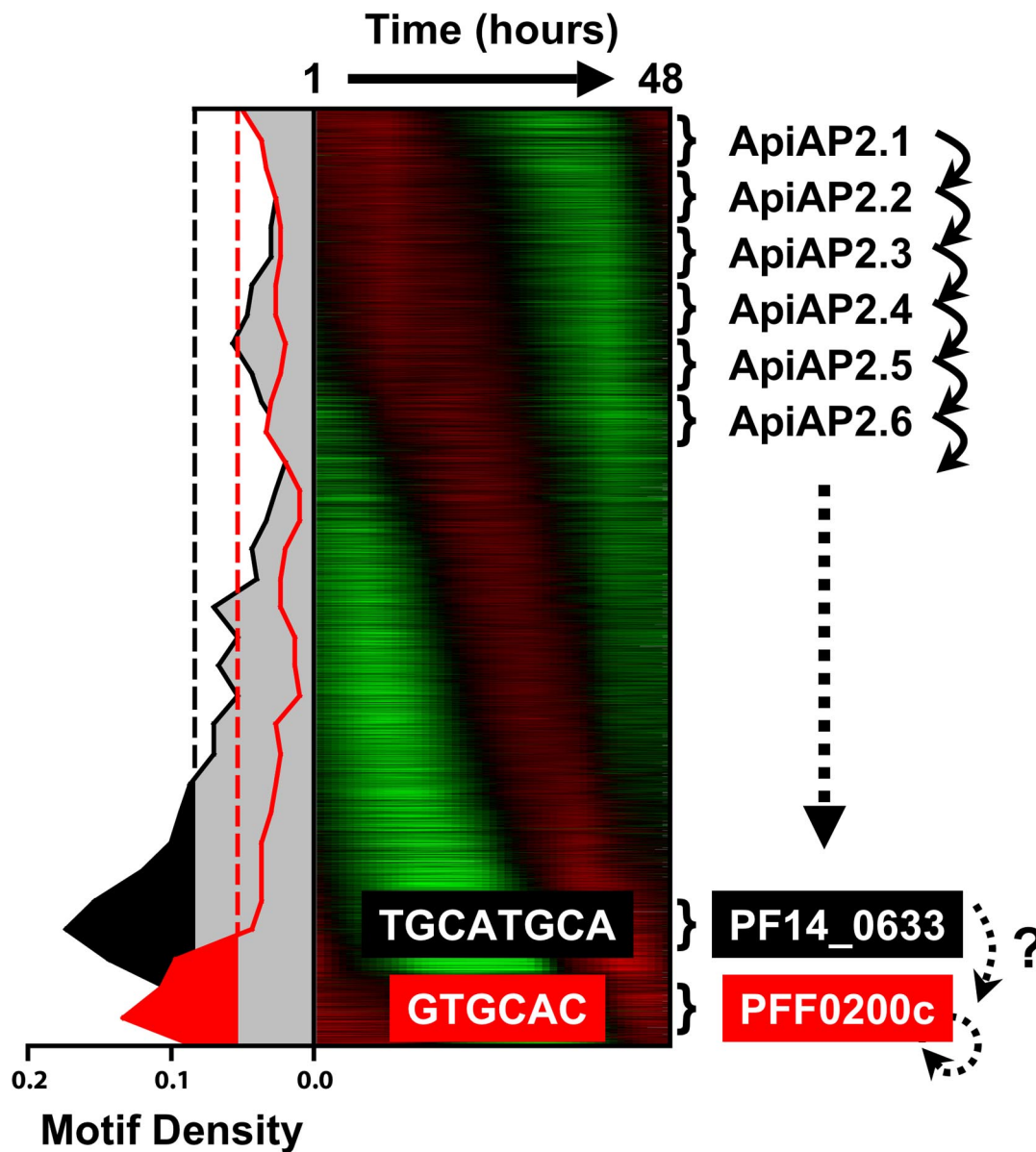


Fig. S3. Model of an ApiAP2 regulatory network throughout the asexual developmental stage of *Plasmodium* development. The 48-hour gene expression profiles are from a previous microarray study by Bozdech *et al.* [Bozdech Z, *et al.* (2003) The transcriptome of the intraerythrocytic developmental cycle of *Plasmodium falciparum*. *PLoS Biol* 1:E5]. The predicted motif density is shown on the left for the two motifs identified in this study. Motif density was calculated as the fraction of genes in a window of 300 with an instance of the given motif. In black is the enrichment for the TGCATGCA motif recognized by PF14.0633, and in red the GTGCAC motif from PFF0200c. The dashed lines represent the significance cutoff (P value = 0.05) and were calculated using standard random permutation analysis repeated 10,000 times and calculating the densities at the 95th percentile. Values below the cutoff are shown in gray. On the right, we propose a model for a cascade of ApiAP2 regulators that includes PF14.0633 potentially regulating PFF0200c as well as autoregulation by PFF0200c.

Table S2. Nuclear localization signals predicted by PredictNLS for ApiAP2 proteins

Gene IDs	Predicted NLS amino acid sequence
MAL8P1.153	KKKKKRKKKK
PF11.0404	KKKINTSEAYHEGKMNKKHKKKK
PF13.0026	RKKKRKEERIVNTGSAKRLEFFYPKKK
PF13.0235	KKKIMNNHHNNKNNKKKKK
PF14.0079	KKKKNQSYYQHNICDDHKSLYDDVKKK
PF14.0271	KKSRSKN
PF14.0471	KRRRNK
PF14.0533	KRKKPNHSKTNDNDQTEIYKTKKMNNK
PFD0200c	KKKKKKKKK
PFD0985w	KRRKQVDVVGDSGTQALKRSKRSKNSKYK
PFF0200c	KHKKKNIHDNNRKK
PFF0670w	KRRRTH
PFF1100c	KKRKSNEEEEEKKNMDLPCCDKDNYIINKLKKNHK
PFL1075w	KKRRNIDEVRKNDNQEKKKKRRK
PFL1085w	KRKYFDDCNMKDEHKKEGKNKQKNKNNKNNKNNK
PFL1900w	KKKKNEDNEYINYKEQNNDERKKKMKDNNKGNKVDKKKK

See Cokol M, Nair R, Rost B (2000) Finding nuclear localization signals. *EMBO Rep* 1: 411–415.

Other Supporting Information Files

[Dataset S1 \(XLS\)](#)

[Dataset S2 \(XLS\)](#)